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DanielBiostatistics10th-package
Functions for Wayne W. Daniel's Biostatistics (Tenth Edition)

Description

Functions and examples to accompany Wayne W. Daniel's *Biostatistics: A Foundation for Analysis in the Health Sciences*, Tenth Edition, Wiley, ISBN: 978-1-119-62550-6.

<https://www.wiley.com/en-us/Biostatistics:+A+Foundation+for+Analysis+in+the+Health+Sciences,+10th+Edition-p-9781119625506>

Data sets from 10th edition <https://bcs.wiley.com/he-bcs/Books?action=resource&bcsId=7849&itemId=1118302796&resourceId=30373>.

Resources from 11th edition <https://bcs.wiley.com/he-bcs/Books?action=index&bcsId=11491&itemId=1119496578>, with errata of data.

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`addProbs`*Conditional and/or Marginal Probabilities*

Description

Add conditional and/or marginal probabilities to a two-way contingency table.

Usage

```
addProbs(A, margin = seq_len(nd), fmt = "%d (%.1f%)")
```

Arguments

`A` [matrix](#) of [typeof integer](#), two-dimensional contingency table. See [addmargins](#)

`margin` [integer](#) scalar or [vector](#), see [addmargins](#)

`fmt` [character](#) scalar, C-style string format with a `%d` and an `%f%` for the counts and proportions (order enforced).

Details

Function [addProbs](#) provides the joint, marginal (using `margin = 1:2`) and conditional (using `margin = 1L` or `margin = 2L`) probabilities of a two-dimensional contingency table.

Value

Function [addProbs](#) returns an 'addProbs' object, which inherits from [table](#) and [noquote](#).

Note

[margin.table](#) (which is to be renamed as [marginSums](#)) is much slower than [colSums](#).

The use of argument `margin` is the same as [addmargins](#), and different from [proportions!](#)

See Also

[rowSums](#) [colSums](#) [proportions](#)

Examples

```
addProbs(table(warpbreaks$tension))

storage.mode(VADeaths) = 'integer'
addProbs(VADeaths)
addProbs(VADeaths, margin = 1L)
rowSums(proportions(VADeaths, margin = 1L))
addmargins(VADeaths, margin = 1L)
```

Description

Functions and examples for Chapter 1, *Introduction to Biostatistics*.

Usage

```
sampleRow(x, size, replace = FALSE, prob = NULL)
```

Arguments

x	a data.frame
size	positive integer scalar, number of rows to be selected
replace	logical scalar, whether sampling should be with replacement (default FALSE)
prob	numeric vector of probability weights for each row of input x being sampled. Default NULL indicates simple random sampling

Value

Function [sampleRow](#) returns a [data.frame](#), a simple random sample from the input.

See Also

[sample.int](#)

Examples

```
library(DanielBiostatistics10th)
# To run a line of code, use shortcut
# Command + Enter: Mac and RStudio Cloud
# Control + Enter: Windows, Mac and RStudio Cloud
# To clear the console
# Control + L: Windows, Mac and RStudio Cloud

# Example 1.4.1; Page 8 (10th ed), Page 7 (11th ed)
class(EXA_C01_S04_01) # `EXA_C01_S04_01` is a 'data.frame' (a specific class defined in R)
dim(EXA_C01_S04_01) # dimension, number-row and number-column
head(EXA_C01_S04_01, n = 8L) # first `n` rows of a 'data.frame'
names(EXA_C01_S04_01) # column names of a 'data.frame'
EXA_C01_S04_01$AGE # use `$` to obtain one column from a 'data.frame'
sampleRow(EXA_C01_S04_01, size = 10L, replace = FALSE) # to answer Example 1.4.1

# Example 1.4.2; Page 11 (10th ed), Page 10 (11th ed)
EXA_C01_S04_01[seq.int(from = 4L, to = 166L, by = 18L), ]
```

Description

Functions and examples for Chapter 2, *Descriptive Statistics*.

Usage

```
print_stats(x, na.rm = TRUE)
```

```
print_freqs(x, breaks, include.lowest = TRUE, right = TRUE)
```

Arguments

x	numeric vector, the observations. In function print_freqs , this argument can also be a factor
na.rm	logical scalar, whether to remove the missing observations (default TRUE)
breaks	numeric vector, see cut.default
include.lowest	logical scalar, default TRUE. See cut.default
right	logical scalar, see cut.default

Details

Function [print_freqs](#) prints the (relative) frequencies and cumulative (relative) frequencies, from a numeric input vector, specified interval breaks as well as open/close status of the ends of the intervals.

Function [print_stats](#) prints the simple statistics of the input observations, such as sample size, mean, median, (smallest) mode, variance, standard deviation, coefficient of variation (if all observations are non-negative), quartiles, inter-quartile range (IQR), range, skewness and kurtosis. A histogram is also printed.

Value

Function [print_freqs](#) returns a [freqs](#) object, for which a [show](#) method is defined.

Function [print_stats](#) does not have a returned value.

See Also

[cut.default](#) [table](#) [cumsum](#) [mean.default](#) [median.default](#) [Mode](#) [var](#) [sd](#) [quantile](#) [skewness](#) [kurtosis](#)

Examples

```

library(DanielBiostatistics10th)

# Example 2.2.1; Page 20 (10th ed), Page 19 (11th ed)
head(EXA_C01_S04_01)
class(age <- EXA_C01_S04_01$AGE) # 'integer'
sort(age) # Table 2.2.1

# Example 2.3.1; Page 23 (10th ed); Page 22 (11th ed)
(r231 = print_freqs(age, breaks = seq.int(from=30, to=90, by=10), right = FALSE)) # Table 2.3.2
# The open/close of interval ends is determined by textbook using 30-39, 40-49, etc.

# Example 2.4.1-2.4.6; Page 38-42 (10th ed); Page 30-34 (11th ed)
# Example 2.5.1-2.5.3; Page 44-46 (10th ed); Page 35-41 (11th ed)
print_stats(age)

# Example 2.5.4 (omitted); Page 49 (10th ed), Page 41 (11th ed)

# Example 2.5.5; Page 50 (10th ed)
head(EXA_C02_S05_05)
boxplot(EXA_C02_S05_05$GRF, main = c('Example 2.5.5 (10th ed)'))
print_stats(EXA_C02_S05_05$GRF)
print_freqs(EXA_C02_S05_05$GRF, breaks = seq.int(10, 45, by = 5))

```

Description

Examples in Chapter 3, *Some Basic Probability Concepts*.

Value

No function defined for Chapter 3.

Examples

```

library(DanielBiostatistics10th)

# Example 3.4.1-3.4.9; Page 69-75 (10th ed), Page 61-67 (11th ed)
(d341 = matrix(c(28L, 19L, 41L, 53L, 35L, 38L, 44L, 60L), ncol = 2L, dimnames = list(
  FamilyHx = c('none', 'Bipolar', 'Unipolar', 'UniBipolar'), Onset = c('Early', 'Late'))))
class(d341) # 'matrix', i.e., a two-dimensional 'array'
addProbs(d341)
addProbs(d341, margin = 2L)
addProbs(d341, margin = 1L)

# Example 3.5.1; Page 81 (10th ed), Page 72 (11th ed)

```

```
(d351 = matrix(c(436L, 14L, 5L, 495L), nrow = 2L, dimnames = list(
  Test = c('Positive', 'Negative'), Alzheimer = c('Yes', 'No'))))
summary(BooleanTable(t(d351)), prevalence = .113)
```

Description

Examples in Chapter 4, *Probability Distributions*.

Value

No function defined for Chapter 4.

Examples

```
library(DanielBiostatistics10th)

# Example 4.2.1-4.2.7; Page 93-97 (10th ed), Page 81-85 (11th ed)
(d421 = rep(1:8, times = c(62L, 47L, 39L, 39L, 58L, 37L, 4L, 11L)))
print_freqs(d421) # Table 4.2.1, 4.2.2, Table 4.2.3

# Example 4.2.8; Page 98 (10th ed), Page 85 (11th ed)
mean(d421)
sd(d421)
var(d421)

# ?dbinom # 'd' for binomial 'density'; calculate Prob(X = x)
# ?pbinom # 'p' for binomial 'probability'
# `lower.tail = TRUE` (default), calculate Prob(X <= x)
# `lower.tail = FALSE`, calculate Prob(X > x)

# Example 4.3.1; Page 99 (10th ed)
dbinom(x = 3L, size = 5L, prob = .858)
# Example 4.3.1; Page 87 (11th ed)
dbinom(x = 3L, size = 5L, prob = .899)

# Example 4.3.2; Page 103 (10th ed), Page 90 (11th ed)
dbinom(x = 4L, size = 10L, prob = .14)

# Example 4.3.3; Page 103 (10th ed), Page 91 (11th ed)
(pL = pbinom(q = 5L, size = 25L, prob = .1, lower.tail = TRUE)) # (a) P(X <= 5L)
(pU = pbinom(q = 5L, size = 25L, prob = .1, lower.tail = FALSE)) # (b) P(X > 5L)
pL + pU # = 1

# Example 4.3.4; Page 105 (10th ed), Page 92 (11th ed)
dbinom(x = 7L, size = 12L, prob = .55)
pbinom(q = 5L, size = 12L, prob = .55)
```

```

pbinom(q = 7L, size = 12L, prob = .55, lower.tail = FALSE)

# Example 4.4.1; Page 110 (10th ed), Page 97 (11th ed)
dpois(x = 3L, lambda = 12)

# Example 4.4.2; Page 110 (10th ed), Page 98 (11th ed)
ppois(q = 2L, lambda = 12, lower.tail = FALSE)

# Example 4.4.3; Page 110 (10th ed), Page 98 (11th ed)
ppois(q = 1L, lambda = 2)

# Example 4.4.4; Page 111 (10th ed), Page 98 (11th ed)
dpois(x = 3L, lambda = 2)

# Example 4.4.5; Page 112 (10th ed), Page 98 (11th ed)
ppois(q = 5L, lambda = 2, lower.tail = FALSE)

# Example 4.6.1; Page 119 (10th ed), Page 106 (11th ed)
pnorm(q = 2)

# Example 4.6.2; Page 120 (10th ed), Page 106 (11th ed)
pnorm(2.55) - pnorm(-2.55)
1 - 2 * pnorm(-2.55) # alternative solution

# Example 4.6.3; Page 121 (10th ed), Page 107 (11th ed)
pnorm(1.53) - pnorm(-2.74)

# Example 4.6.4; Page 121 (10th ed), Page 107 (11th ed)
pnorm(2.71, lower.tail = FALSE)

# Example 4.6.5; Page 122 (10th ed), Page 107 (11th ed)
pnorm(2.45) - pnorm(.84)

# Example 4.7.1; Page 122 (10th ed), Page 109 (11th ed)
pnorm(q = 3, mean = 5.4, sd = 1.3)
pnorm(q = (3-5.4)/1.3) # manual solution

# Example 4.7.2; Page 125 (10th ed), Page 111 (11th ed)
pnorm(q = 649, mean = 491, sd = 119) - pnorm(q = 292, mean = 491, sd = 119)

# Example 4.7.3; Page 122 (10th ed), Page 111 (11th ed)
1e4L * pnorm(q = 8.5, mean = 5.4, sd = 1.3, lower.tail = FALSE)

```

Description

Functions for Chapter 5, *Some Important Sampling Distributions*, Chapter 6, *Estimation* and Chapter 7, *Hypothesis Testing*.

Usage

```

aggregated_z(
  xbar,
  n,
  sd,
  null.value,
  alternative = c("two.sided", "less", "greater"),
  conf.level = 0.95,
  ...
)

```

```

aggregated_t(
  xbar,
  xsd,
  n,
  null.value,
  var.equal = FALSE,
  alternative = c("two.sided", "less", "greater"),
  conf.level = 0.95,
  ...
)

```

```

prop_CLT(
  x,
  n,
  obs,
  xbar = x/n,
  null.value,
  alternative = c("two.sided", "less", "greater"),
  conf.level = 0.95,
  ...
)

```

```

aggregated_var(
  xsd,
  n,
  null.value,
  alternative = c("two.sided", "less", "greater"),
  conf.level = 0.95,
  ...
)

```

Arguments

xbar **numeric** scalar or **length-2 vector**. Sample mean(s) for **numeric** variable(s) \bar{x} or (\bar{x}_1, \bar{x}_2) . Sample proportion(s) for binary (i.e., **logical**) variable(s) \hat{p} or (\hat{p}_1, \hat{p}_2) . In the case of two-sample tests, this could also be a **numeric** scalar indicating the difference in sample means $\bar{x}_1 - \bar{x}_2$ or sample proportions $\hat{p}_1 - \hat{p}_2$

<code>n</code>	integer scalar n or length-2 vector (n_1, n_2) , sample size(s)
<code>sd</code>	numeric scalar σ or length-2 vector (σ_1, σ_2) , population standard deviation(s)
<code>null.value</code>	(optional) numeric scalar or length-2 vector . Null value(s) of the population mean(s) $(\mu_0, (\mu_{10}, \mu_{20}),$ or $\mu_{10} - \mu_{20})$ for functions <code>aggregated_z</code> and <code>aggregated_t</code> . Null value(s) of the population proportion(s) $(p_0, (p_{10}, p_{20}),$ or $p_{10} - p_{20})$ for <code>prop_CLT</code> . Null value(s) of the population variance(s) (ratio) $(\sigma_0^2, (\sigma_{10}^2, \sigma_{20}^2),$ or $\sigma_{10}^2/\sigma_{20}^2)$ for function <code>aggregated_var</code> . If missing, only the confidence intervals will be computed.
<code>alternative</code>	character scalar, alternative hypothesis, either 'two.sided' (default), 'greater' or 'less'
<code>conf.level</code>	numeric scalar $(1 - \alpha)$, confidence level, default 0.95
<code>...</code>	potential arguments, not in use currently
<code>xsd</code>	numeric scalar s or length-2 vector (s_1, s_2) , sample standard deviation(s)
<code>var.equal</code>	logical scalar, whether to treat the two population variances as being equal (default FALSE) in function <code>aggregated_t</code>
<code>x</code>	integer scalar or length-2 vector , number of positive count(s) of binary (i.e., logical) variable(s)
<code>obs</code>	vector , observations, currently used only in one-sample z -test on proportion <code>prop_CLT</code>

Details

Function `aggregated_z` performs one- or two-sample z -test using the aggregated statistics of sample mean(s) and sample size(s) when `null.value` is provided. Otherwise, only the confidence interval based on z -distribution is computed.

Function `aggregated_t` performs one- or two-sample t -test using the aggregated statistics of sample mean(s), sample standard deviation(s) and sample size(s) when `null.value` is provided. Otherwise, only the confidence interval based on t -distribution is computed.

Function `prop_CLT` performs one- or two-sample z -test on proportion(s), using Central Limit Theorem when `null.value` is provided. Otherwise, only the confidence interval based on z -distribution is computed.

Function `aggregated_var` performs one-sample χ^2 -test on variance, or two-sample F -test on variances, using the aggregated statistics of sample standard deviation(s) and sample size(s) when `null.value` is provided. Otherwise, only the confidence interval based on χ^2 - or F -distribution is computed.

Value

Function `aggregated_z` returns an 'htest' object when `null.value` is provided, otherwise returns a length-two **numeric** vector.

Function `aggregated_t` returns an `htest` object when `null.value` is provided, otherwise returns a length-two **numeric** vector.

Function `prop_CLT` returns an `htest` object when `null.value` is provided, otherwise returns a length-two **numeric** vector.

Function `aggregated_var` returns an `htest` object when `null.value` is provided, otherwise returns a length-two **numeric** vector.

See Also

[t.test prop.test var.test](#)

Examples

```
library(DanielBiostatistics10th)

# Example 5.3.2; Page 142 (10th ed)
aggregated_z(xbar = 190, sd = 12.7, n = 10L, null.value = 185.6, alternative = 'greater')

# Example 5.3.3;
# Page 143 (10th ed)
pnorm(125, mean = 120, sd = 15/sqrt(50)) - pnorm(115, mean = 120, sd = 15/sqrt(50))
aggregated_z(125, sd = 15, n = 50L, null.value = 120, alternative = 'less')$p.value -
  aggregated_z(115, sd = 15, n = 50L, null.value = 120, alternative = 'less')$p.value
# Page 127 (11th ed)
aggregated_z(110, sd = 20, n = 50L, null.value = 100, alternative = 'less')$p.value -
  aggregated_z(95, sd = 20, n = 50L, null.value = 100, alternative = 'less')$p.value

# Example 5.4.1; Page 145 (10th ed), Page 129 (11th ed)
aggregated_z(xbar = c(92, 105), sd = 20, n = 15L, null.value = 0, alternative = 'less')

# Example 5.4.2; Page 148 (10th ed),
aggregated_z(xbar = 20, sd = c(15, 20), n = c(35L, 40L), null.value = c(45, 30),
  alternative = 'greater')

# Example 5.5.1; Page 150 (10th ed),
prop_CLT(xbar = .4, n = 150L, null.value = .357, alternative = 'greater')

# Example 5.5.2; Page 152 (10th ed),
prop_CLT(xbar = .45, n = 200L, null.value = .51, alternative = 'less')

# Example 5.6.1; Page 155 (10th ed),
prop_CLT(xbar = .1, null.value = c(.28, .21), n = c(100L, 100L), alternative = 'greater')

# Example 5.6.2; Page 155 (10th ed),
prop_CLT(xbar = .05, null.value = c(.34, .26), n = c(250L, 200L), alternative = 'less')

# Example 6.2.1; Page 166 (10th ed), Page 147 (11th ed)
aggregated_z(xbar = 22, n = 10L, sd = sqrt(45))

# Example 6.2.2; Page 168 (10th ed), Page 149 (11th ed)
aggregated_z(xbar = 84.3, n = 15L, sd = sqrt(144), conf.level = .99)

# Example 6.2.3; Page 168 (10th ed), Page 150 (11th ed)
aggregated_z(xbar = 17.2, n = 35L, sd = 8, conf.level = .9)

# Example 6.2.4; Page 169 (10th ed), Page 150 (11th ed)
head(EXA_C06_S02_04)
aggregated_z(xbar = mean(EXA_C06_S02_04$ACTIVITY), n = nrow(EXA_C06_S02_04), sd = sqrt(.36))

# Example 6.3.1; Page 173 (10th ed),
```

```

aggregated_t(xbar = 250.8, xsd = 130.9, n = 19L)

# Example 6.4.1; Page 177 (10th ed),
aggregated_z(xbar = c(4.5, 3.4), sd = sqrt(c(1, 1.5)), n = c(12L, 15L))

# Example 6.4.2; Page 178 (10th ed),
aggregated_z(xbar = c(4.3, 13), sd = c(5.22, 8.97), n = c(328L, 64L), conf.level = .99)

# Example 6.4.3; Page 180 (10th ed),
aggregated_t(xbar = c(4.7, 8.8), xsd = c(9.3, 11.5), n = c(18L, 10L), var.equal = TRUE)

# Example 6.4.4; Page 181 (10th ed),
aggregated_t(xbar = c(4.7, 8.8), xsd = c(9.3, 11.5), n = c(18L, 10L))
# Welch slightly different from Cochran; textbook explained on Page 182

# Example 6.5.1; Page 185 (10th ed),
prop_CLT(xbar = .18, n = 1220L)

# Example 6.6.1; Page 187 (10th ed),
prop_CLT(x = c(31L, 53L), n = c(68L, 255L), conf.level = .99)

# Example 6.7.1; Page 190 (10th ed),
(n_671 = uniroot(f = function(n, sd, level = .95) {
  qnorm(1-(1-level)/2) * sd/sqrt(n) - 5 # half-width of CI <= 5 grams
}, interval = c(0, 2e2), sd = 20)$root)
ceiling(n_671)

# Example 6.8.1; Page 192 (10th ed),
(n_681 = uniroot(f = function(n, p, level = .95) {
  qnorm(1-(1-level)/2) * sqrt(p*(1-p)/n) - .05
}, interval = c(0, 1e3), p = .35)$root)
ceiling(n_681)

# Example 6.9.1; Page 196 (10th ed),
d691 = c(9.7, 12.3, 11.2, 5.1, 24.8, 14.8, 17.7)
sqrt(aggregated_var(xsd = sd(d691), n = length(d691)))

# Example 6.10.1; Page 200 (10th ed),
aggregated_var(xsd = c(8.1, 5.9), n = c(16L, 4L))

# Example 7.2.1; Page 222 (10th ed); Page 201 (11th ed)
aggregated_z(xbar = 27, sd = sqrt(20), n = 10L, null.value = 30)

# Example 7.2.2; Page 226 (10th ed); Page 204 (11th ed)
aggregated_z(xbar = 27, sd = sqrt(20), n = 10L, null.value = 30, alternative = 'less')

# Example 7.2.3; Page 228 (10th ed); Page 206 (11th ed)
head(EXA_C07_S02_03)
t.test(EXA_C07_S02_03$DAYS, mu = 15)

# Example 7.2.4; Page 231 (10th ed); Page 209 (11th ed)
aggregated_z(xbar = 146, sd = 27, n = 157L, null.value = 140, alternative = 'greater')

```

```

# Example 7.2.5; Page 232 (10th ed); Page 210 (11th ed)
d725 = c(33.38, 32.15, 34.34, 33.95, 33.46, 34.13, 33.99, 34.10, 33.85,
  34.23, 34.45, 34.19, 33.97, 32.73, 34.05)
t.test(d725, mu = 34.5)

# Example 7.3.1; Page 237 (10th ed), Page 213 (11th ed)
aggregated_z(xbar = c(4.5, 3.4), sd = sqrt(c(1, 1.5)), n = c(12L, 15L), null.value = 0)

# Example 7.3.2; Page 239 (10th ed), Page 215 (11th ed)
head(EXA_C07_S03_02)
with(EXA_C07_S03_02, t.test(x = CONTROL, y = SCI, alternative = 'less', var.equal = TRUE))

# Example 7.3.3; Page 240 (10th ed), Page 217 (11th ed)
aggregated_t(xbar = c(19.16, 9.53), xsd = c(5.29, 2.69), n = c(15L, 30L), null.value = 0)

# Example 7.3.4; Page 242 (10th ed), Page 219 (11th ed)
aggregated_z(xbar = c(59.01, 46.61), sd = c(44.89, 34.85), n = c(53L, 54L), null.value = 0,
  alternative = 'greater')

# Example 7.4.1; Page 251 (10th ed), Page 226 (11th ed)
head(EXA_C07_S04_01)
with(EXA_C07_S04_01, t.test(x = POSTOP, y = PREOP, alternative = 'greater', paired = TRUE))

# Example 7.5.1; Page 258 (10th ed), Page 232 (11th ed)
prop_CLT(x = 24L, n = 301L, null.value = .063, alternative = 'greater')

# Example 7.6.1; Page 261 (10th ed), Page 235 (11th ed)
prop_CLT(x = c(24L, 11L), n = c(44L, 29L), null.value = 0, alternative = 'greater')

# Example 7.7.1; Page 264 (10th ed), Page 238 (11th ed)
head(EXA_C07_S07_01)
aggregated_var(xsd = sd(EXA_C07_S07_01$mass), n = 16L, null.value = 600)

# Example 7.8.1; Page 268 (10th ed), Page 242 (11th ed)
aggregated_var(xsd = c(30.62, 11.37), n = 6L, null.value = 1, alternative = 'greater')

# Example 7.8.2; Page 270 (10th ed),
with(EXA_C07_S03_02, var.test(x = CONTROL, y = SCI))

```

Description

Functions for Chapter 7, *Hypothesis Testing*.

Usage

```
power_z(
  x,
```

```

null.value,
sd,
n,
alternative = c("two.sided", "less", "greater"),
sig.level = 0.05
)

```

Arguments

<code>x</code>	numeric vector , mean parameter(s) μ_1 in the alternative hypothesis
<code>null.value</code>	numeric scalar , mean parameter μ_0 in the null hypothesis
<code>sd</code>	numeric scalar , population standard deviation σ
<code>n</code>	integer scalar , sample size n
<code>alternative</code>	character scalar , alternative hypothesis, either 'two.sided' (default), 'greater' or 'less'
<code>sig.level</code>	numeric scalar , significance level (i.e., Type-I-error rate), default .05

Details

Function `power_z` calculates the powers at each element of the alternative parameters μ_1 , for one-sample z -test

- $H_0 : \mu = \mu_0$ vs. $H_A : \mu \neq \mu_0$, if `alternative = 'two.sided'`
- $H_0 : \mu \leq \mu_0$ vs. $H_A : \mu > \mu_0$, if `alternative = 'greater'`
- $H_0 : \mu \geq \mu_0$ vs. $H_A : \mu < \mu_0$, if `alternative = 'less'`

Value

Function `power_z` returns a 'power_z' object, which inherits from 'power.htest' class.

See Also

[power.t.test](#)

Examples

```

library(DanielBiostatistics10th)

# Example 7.9.1; Page 272 (10th ed), Page 245 (11th ed)
(p791 = power_z(seq.int(from = 16, to = 19, by = .5), null.value = 17.5, sd = 3.6, n = 100L))
# Table 7.9.1

# Example 7.9.2; Page 276 (10th ed), Page 248 (11th ed)
(p792 = power_z(seq.int(from = 50, to = 70, by = 5), null.value = 65, sd = 15, n = 20L,
  sig.level = .01, alternative = 'less'))

# Example 7.10.1; Page 278 (10th ed),
(n_d7101 <- uniroot(f = function(x) {
  power_z(55, null.value = 65, sd = 15, n = x, sig.level = .01, alternative = 'less')$power - .95

```

```

}, interval = c(0, 50))$root)
power_z(55, null.value = 65, sd = 15, n = ceiling(n_d7101), sig.level = .01, alternative = 'less')

```

Description

Examples for Chapter 8, *Analysis of Variance*.

Value

No function defined for Chapter 8.

Examples

```

library(DanielBiostatistics10th)

# Example 8.2.1; Page 318 (10th ed), Page 280 (11th ed)
head(EXA_C08_S02_01)
boxplot(selenium ~ type, data = EXA_C08_S02_01, main = 'Figure 8.2.7')
(aov_d821 = aov(selenium ~ type, data = EXA_C08_S02_01))
# ?stats::aov # analysis-of-variance model
anova(aov_d821)
# ?stats::anova # ANOVA table

# Example 8.2.2; Page 325 (10th ed), Page 286 (11th ed)
(tukey_d822 <- TukeyHSD(aov_d821, conf.level = 0.95)) # Figure 8.2.8
plot(tukey_d822)

# Example 8.3.1; Page 339 (10th ed), Page 298 (11th ed)
head(EXA_C08_S03_01)
head(d831 <- within(EXA_C08_S03_01, expr = {
  ageGroup = structure(ageGroup, levels = c('<20', '20s', '30s', '40s', '>50'), class = 'factor')
}))
(aov_831 = aov(time ~ method + ageGroup, data = d831))
anova(aov_831)

# Example 8.4.1; Page 348 (10th ed), Page 307 (11th ed)
head(EXA_C08_S04_01)
head(d841 <- within(EXA_C08_S04_01, expr = {
  SUBJ = factor(SUBJ)
  TIME = structure(TIME, levels = c('Baseline', '1-Mon', '3-Mon', '6-Mon'), class = 'factor')
}))
(aov_841 = aov(FUNC ~ SUBJ + TIME, data = d841))
anova(aov_841)

# Example 8.4.2; Page 352 (10th ed), Page 310 (11th ed)
# (optional; out of the scope of this course)
head(EXA_C08_S04_02)

```

```

names(EXA_C08_S04_02)[3:6] = c('baseline', '2wk', '4wk', '6wk')
head(d842a <- within(EXA_C08_S04_02, expr = {
  subject = factor(subject)
  treatment = structure(treatment, levels = c('placebo', 'aloe_juice'), class = 'factor')
}))
head(d842b <- reshape2::melt(d842a, id.vars = c('subject', 'treatment'),
  variable.name = 'time', value.name = 'OralScores'))

# Hypothesis:
# Main effect of 'treatment';
# Main effect of 'time';
# Interaction between 'treatment' and 'time'
(aov_842 = aov(OralScores ~ treatment * time + Error(subject), data = d842b))
class(aov_842)
summary(aov_842)
# Section 'Error: subject' in R output
# .. is Figure 8.4.4 'Tests of Between-Subjects Effects' (without the row of 'Intercept')
# .. 'treatment' row: effect of treatment at **baseline** (i.e. reference time),
# ... degree-of-freedom (dof) = 2-1 = 1
# .. 'Residuals' row: residual at baseline, dof = (25-1) - (2-1) = 23
# .. It's important to note that 'treatment' is a **between-subject factor**.
# Section 'Error: Within' in R output
# .. is Figure 8.4.4 'Tests of Within-Subjects Effects'
# .. 'time' row: effect of time within subject for placebo (i.e. reference treatment), dof = 4-1 = 3
# .. 'treatment:time' row: interaction of treatment and time, dof = (2-1)*(4-1) = 3
# .. 'Residuals' row: residual at 2wk, 4wk and 6wk, dof = (4-1)*23 = 69
# ... [(4-1) timepoints, 23 dof at each timepoints]
# Analysis Interpretation
# .. No signif. diff. detected between placebo vs. aloe at baseline (p = .815)
# .. No signif. diff. detected in the trends over time between placebo vs. aloe (p = .974)
# .. Signif. diff. detected among the four timepoints, for either placebo or aloe pts (p = 3e-7)
# R code below creates an equivalent ANOVA model
anova(aov(OralScores ~ treatment * time + subject, data = d842b))
# .. 'subject' is considered as a block factor

# Example 8.5.2; Page 364 (10th ed), Page 321 (11th ed)
head(EXA_C08_S05_02)
head(d852 <- within(EXA_C08_S05_02, expr = {
  A = structure(A, levels = c('Cardiac', 'Cancer', 'CVA', 'Tuberculosis'), class = 'factor')
  B = structure(B, levels = c('20s', '30s', '40s', '50+'), class = 'factor')
}))
(aov_852 = aov(HOME ~ A * B, data = d852))
anova(aov_852)
summary(lm(HOME ~ A * B, data = d852))
# produces alpha, beta and (alpha beta)'s in the formulation

```

Description

Examples in Chapter 9, *Simple Linear Regression and Correlation*.

Value

No function defined for Chapter 9.

Examples

```
library(DanielBiostatistics10th)

# Example 9.3.1; Page 417 (10th ed), Page 358 (11th ed)
head(EXA_C09_S03_01)
names(EXA_C09_S03_01)[2:3] = c('Waist', 'AT')
plot(AT ~ Waist, data = EXA_C09_S03_01,
      xlab = 'Waist circumference (cm), X',
      ylab = 'Deep abdominal AT area (cm2), Y',
      main = 'Figure 9.3.1')
m931 = lm(AT ~ Waist, data = EXA_C09_S03_01)
plot(AT ~ Waist, data = EXA_C09_S03_01,
      xlab = 'Waist circumference (cm), X',
      ylab = 'Deep abdominal AT area (cm2), Y',
      main = 'Figure 9.3.3'); abline(m931, col = 'red', lty = 2)

# Example 9.4.1; Page 432 (10th ed), Page 372 (11th ed)
anova(m931) # Table 9.4.1. Page 434 (10th ed)

# Example 9.4.2; Page 436 (10th ed), Page 375 (11th ed)
summary(m931)
confint(m931) # Page 438 (10th ed)

# (omitted) Example 9.4.3; Page 376 (11th ed)

# (optional)
# Example 9.4.3; Page 440 (10th ed)
# Example 9.4.4; Page 379 (11th ed)
plot(m931, which = 1, main = 'Figure 9.4.8 (10th) or 9.4.9 (11th)')

# Example 9.7.1; Page 447 (10th ed), Page 386 (11th ed)
head(EXA_C09_S07_01)
plot(CV ~ HEIGHT, data = EXA_C09_S07_01,
      xlab = 'Height (cm)',
      ylab = 'Cv (units)',
      main = 'Figure 9.7.2')
m971 = lm(CV ~ HEIGHT, data = EXA_C09_S07_01)
summary(m971); anova(m971) # Figure 9.7.3; Page 450 (10th ed)

# Example 9.7.2; Page 452 (10th ed), Page 390 (11th ed)
cor(EXA_C09_S07_01);
cor.test(~ CV + HEIGHT, data = EXA_C09_S07_01) # Figure 9.7.4, 9.7.5

# Page 453, When the Hypothesized rho Is a Nonzero Value
# R does not have a function to do this
```

Description

Examples for Chapter 10, *Multiple Regression and Correlation*.

Value

No function defined for Chapter 10.

Examples

```
library(DanielBiostatistics10th)

# Example 10.3.1; Page 493 (10th ed), Page 419 (11th ed)
head(EXA_C10_S03_01)
pairs(EXA_C10_S03_01, main = 'Figure 10.3.1')
m1031 = lm(CDA ~ AGE + EDLEVEL, data = EXA_C10_S03_01)
summary(m1031)

# Example 10.4.1; Page 502 (10th ed), Page 428 (11th ed)
# .. see 'Multiple R-squared' (not 'Adjusted R-squared')

# Example 10.4.2; Page 504 (10th ed), Page 429 (11th ed)
# .. see 'F-statistic'

# Example 10.4.3; Page 505 (10th ed), Page 430 (11th ed)
# .. see 'Coefficients:'

# confidence interval for beta's; Page 506 (10th ed), Page 431 (11th ed)
confint(m1031)

# Example 10.5.1; Page 509 (10th ed), Page 434 (11th ed)
(newd_1031 = data.frame(AGE = 68, EDLEVEL = 12))
predict(m1031, newdata = newd_1031, interval = 'prediction')
predict(m1031, newdata = newd_1031, interval = 'confidence')

# Example 10.6.1; Page 511 (10th ed), Page 436 (11th ed)
head(EXA_C10_S06_01)
pairs(EXA_C10_S06_01, main = 'Scatter Plot Matrix, Example 10.6.1')
m1061 = lm(W ~ P + S, data = EXA_C10_S06_01)
summary(m1061)
confint(m1061)
anova(m1061)

# (optional)
# Example 10.6.2; Page 515 (10th ed), Page 440 (11th ed)
psych::partial.r(EXA_C10_S06_01, x = 2:3, y = 1L)
```

Description

Examples in Chapter 11, *Regression Analysis: Some Additional Techniques*.

Value

No function defined for Chapter 11.

Examples

```
library(DanielBiostatistics10th)

# Example 11.1.1; Page 540,
head(EXA_C11_S01_01)
head(log(EXA_C11_S01_01$conc, base = 10))
head(EXA_C11_S01_01$logConc)

# Example 11.1.2; Page 542,
head(EXA_C11_S01_02)
cor.test(~ sbp + weight, data = EXA_C11_S01_02)
cor.test(~ sbp + bmi, data = EXA_C11_S01_02)

# Example 11.2.1; Page 545,
head(EXA_C11_S02_01)
d1121 = within(EXA_C11_S02_01, expr = {
  SMOKE = as.logical(SMOKE)
})
xlab1121 = 'Length of gestation (weeks)'; ylab1121 = 'Birth weight (grams)'
car::scatterplot(GRAMS ~ WEEKS | SMOKE, data = d1121, regLine = FALSE, smooth = FALSE,
  xlab = xlab1121, ylab = ylab1121, main = 'Figure 11.2.1')
m1121 = lm(GRAMS ~ WEEKS + SMOKE, data = d1121)
summary(m1121) # Figure 11.2.2
confint(m1121)
car::scatterplot(GRAMS ~ WEEKS | SMOKE, data = d1121, regLine = FALSE, smooth = FALSE,
  xlab = xlab1121, ylab = ylab1121, main = 'Figure 11.2.3')
(cf1121 = m1121$coefficients)
abline(a = cf1121[1L], b = cf1121[2L], col = 'blue') # regression line for non-smoking mothers
abline(a = cf1121[1L] + cf1121[3L], b = cf1121[2L], col = 'magenta')

# Example 11.2.3; Page 551,
d1123 = within(EXA_C11_S02_03, expr = {
  METHOD = factor(METHOD, levels = c('C', 'A', 'B')) # textbook designated 'C' as reference level
})
summary(m1123 <- lm(EFFECT ~ AGE * METHOD, data = d1123)) # Figure 11.2.5
confint(m1123)
car::scatterplot(EFFECT ~ AGE | METHOD, data = d1123, smooth = FALSE,
  xlab = 'Age', ylab = 'Treatment effectiveness', main = 'Figure 11.2.6')
```

```

# Example 11.3.1; Page 561,
head(EXA_C11_S03_01)
names(EXA_C11_S03_01) = c('JOBPER', 'ASRV', 'ENTH', 'AMB', 'COMM', 'PROB', 'INIT')
summary(m1131_raw <- lm(JOBPER ~ ASRV + ENTH + AMB + COMM + PROB + INIT, data = EXA_C11_S03_01))
# summary(m1131 <- MASS::stepAIC(m1131_raw, direction = 'backward'))
# the stepwise selection criterion used in MINITAB is not necessarily AIC

# Example 11.4.1; Page 572,
addmargins(d1141 <- array(c(92L, 21L, 15L, 20L), dim = c(2L, 2L), dimnames = list(
  OCAD = c('Present', 'Absent'), Sex = c('Male', 'Female')))) # Table 11.4.2
(d1141a = within(as.data.frame(table(as.table(d1141))), expr = {
  OCAD = (OCAD == 'Present')
  Sex = factor(Sex, levels = c('Female', 'Male'))
}))
m1141 = glm(OCAD ~ Sex, family = binomial, weights = Freq, data = d1141a)
summary(m1141) # Figure 11.4.1
exp(m1141$coefficients[2L]) # exp(beta_M)
exp(confint(m1141)) # confidence interval of exp(beta)
predict(m1141, newdata = data.frame(Sex = setNames(nm = c('Male', 'Female'))), type = 'response')

# Example 11.4.2; Page 573,
head(EXA_C11_S04_02)
summary(m1142 <- glm(ATT ~ AGE, family = binomial, data = EXA_C11_S04_02)) # Figure 11.4.2
exp(m1142$coefficients[2L])
exp(confint(m1142))
car::Anova(m1142) # Optional

# Example 11.4.3; Page 576,
head(REV_C11_24)
summary(glm(ONSET ~ HIAA + TRYPT, family = binomial, data = REV_C11_24)) # Figure 11.4.4
# Predictor TRYPT should be removed from model due to p-value \approx 1
summary(glm(ONSET ~ HIAA, family = binomial, data = REV_C11_24))

# Example 11.4.4–11.4.5; Page 578–579
DescTools::PseudoR2(m1142, which = 'CoxSnell')
DescTools::PseudoR2(m1142, which = 'Nagelkerke')

```

Description

Functions for Chapter 12, *The Chi-Square Distribution and the Analysis of Frequencies*.

Usage

```
print_OE(0, prob)
```

Arguments

0 **integer** vector, observed counts

prob **numeric** vector, anticipated probability. If missing (default), an uniform distribution across all categories are used.

Value

Function `print_OE` prints a table with observed and expected frequencies, as well as the category-wise χ^2 statistics. A **double** vector of the category-wise χ^2 statistics is returned invisibly.

Examples

```
library(DanielBiostatistics10th)

# Example 12.3.1; Page 605 (10th ed)
d1231_b = c(-Inf, seq.int(from = 125, to = 275, by = 25), Inf)
(d1231 = setNames( # Table 12.3.1
  c(1L, 3L, 8L, 18L, 6L, 4L, 4L, 3L),
  nm = levels(cut(double(), breaks = d1231_b, right = FALSE, include.lowest = TRUE))))
chi1231 = print_OE(d1231, prob = diff.default(pnorm(q = d1231_b, mean = 198.67, sd = 41.31)))
pchisq(sum(chi1231), df = length(d1231) - 3L, lower.tail = FALSE)
# -3L: three restrictions (explained on Page 608)
# (1) making sum(xo) == sum(xe)
# (2) estimating mean
# (3) estimating sd

# Example 12.3.2; Page 609 (10th ed),
# 100 doctors, 25 patients per doctor
d1232 = c(5L, 6L, 8L, 10L, 10L, 15L, 17L, 10L, 10L, 9L, 0L)
o1232 = setNames(c(sum(d1232[1:2]), d1232[-(1:2)]), nm = c('0-1', '2-9', '10 or more'))
(p1232 = sum((0:10) * d1232) / (25 * 100)) # binomial `prob`
chi1232 = print_OE(o1232, prob = c(
  pbinom(1L, size = 25L, prob = p1232),
  dbinom(2:9, size = 25L, prob = p1232),
  pbinom(9, size = 25L, prob = p1232, lower.tail = FALSE)))
pchisq(sum(chi1232), df = length(o1232) - 2L, lower.tail = FALSE)
# -2L: two restrictions (explained on Page 611)
# (1) making sum(o) == sum(e)
# (2) estimating p1232

# Example 12.3.3; Page 611 (10th ed),
d1233 = c(5L, 14L, 15L, 23L, 16L, 9L, 3L, 3L, 1L, 1L, 0L)
o_1233 = setNames(c(d1233[1:8], sum(d1233[-(1:8)])), nm = c(0:7, '8 or more'))
p_1233 = c(dpois(0:7, lambda = 3), # lambda = 3 is provided by the textbook
  ppois(7L, lambda = 3, lower.tail = FALSE))
chi1233 = print_OE(o_1233, prob = p_1233)
pchisq(sum(chi1233), df = length(o_1233) - 1L, lower.tail = FALSE)
# -1L: one restrictions
# (1) making sum(xo) == sum(xe)
chisq.test(o_1233, p = p_1233) # equivalent # warning on any(E < 5)
```

```

# Example 12.3.4; Page 614 (10th ed), Page 531 (11th ed)
d1234 = c('Dec 05' = 62L, 'Jan 06' = 84L, 'Feb 06' = 17L, 'Mar 06' = 16L, 'Apr 06' = 21L)
print_OE(d1234) # Figure 12.3.2
chisq.test(d1234)

# Example 12.3.5; Page 616 (10th ed), Page 533 (11th ed)
d1235 = c(dominant = 43L, heterozygous = 125L, recessive = 32L)
print_OE(d1235, prob = c(1, 2, 1))
chisq.test(d1235, p = c(1, 2, 1), rescale.p = TRUE)

# Example 12.4.1; Page 621 (10th ed)
addmargins(d1241 <- array(c(260L, 15L, 7L, 299L, 41L, 14L), dim = c(3L, 2L), dimnames = list(
  Race = c('White', 'Black', 'Other'), FolicAcid = c('TRUE', 'FALSE'))))
chisq.test(d1241) # ?stats::chisq.test

# Example 12.4.2; Page 626 (10th ed),
addmargins(d1242 <- array(c(131L, 14L, 52L, 36L), dim = c(2L, 2L), dimnames = list(
  Type = c('Faller', 'Non-Faller'), LifestyleChange = c('TRUE', 'FALSE'))))
chisq.test(d1242, correct = FALSE)
chisq.test(d1242, correct = TRUE) # Yates's Correction

# Example 12.5.1; Page 631 (10th ed),
addmargins(d1251 <- array(c(21L, 19L, 75L, 77L), dim = c(2L, 2L), dimnames = list(
  Group = c('Narcoleptic', 'Healthy'), Migraine = c('TRUE', 'FALSE'))))
(chisq_1251 = chisq.test(d1251, correct = FALSE))
if (FALSE) {
  # (optional) using test on two proportions
  # only equivalent for 2*2 contingency table
  (clt_1251 = prop_CLT(x = c(21L, 19L), n = 96L, null.value = 0))
  all.equal.numeric(unname(clt_1251$statistic^2), unname(chisq_1251$statistic))
}

# Example 12.6.1; Page 638 (10th ed),
addmargins(d1262 <- array(c(2L, 8L, 7L, 4L), dim = c(2L, 2L), dimnames = list(
  Group = c('PI Naive', 'PA Experienced'), Regimen2yr = c('TRUE', 'FALSE'))))
fisher.test(d1262)

# Example 12.7.1; Page 644 (10th ed),
addmargins(d1271 <- array(c(22L, 18L, 216L, 199L), dim = c(2L, 2L), dimnames = list(
  Exercising = c('Extreme', 'No'), PretermLabor = c('Cases', 'Noncases'))))
DescTools::RelRisk(d1271, conf.level = .95) # 95% CI for relative risk
fisher.test(d1271) # 95% CI for odds ratio

# Example 12.7.2; Page 647 (10th ed),
addmargins(d1272 <- array(c(64L, 68L, 342L, 3496L), dim = c(2L, 2L), dimnames = list(
  SmkPregnancy = c('Smoked Throughout', 'Never Smoked'), Obesity = c('Cases', 'Noncases'))))
fisher.test(d1272)

# Example 12.7.3-12.7.4; Page 650-652 (10th ed),
(d1273 <- array(c(21L, 16L, 11L, 6L, 50L, 18L, 14L, 6L), dim = c(2L, 2L, 2L), dimnames = list(
  HTN = c('Present', 'Absent'), OCAD = c('Cases', 'Controls'), Age = c('<=55', '>55'))))
addmargins(d1273, margin = 1:2) # Table 12.7.6
mantelhaen.test(d1273)

```

Description

Examples for Chapter 13, *Nonparametric and Distribution-Free Statistics*.

Value

No function defined for Chapter 13.

Examples

```
library(DanielBiostatistics10th)

# Example 13.3.1; Page 673,
d1331 = c(4, 5, 8, 8, 9, 6, 10, 7, 6, 6)
BSDA::SIGN.test(d1331, md = 5)

# Example 13.3.2; Page 677,
head(EXA_C13_S03_02)
with(EXA_C13_S03_02, BSDA::SIGN.test(x = X, y = Y, alternative = 'less'))

# Example 13.4.1; Page 683,
d1341 = c(4.91, 4.10, 6.74, 7.27, 7.42, 7.50, 6.56, 4.64, 5.98, 3.14, 3.23, 5.80, 6.17, 5.39, 5.77)
wilcox.test(d1341, mu = 5.05)

# Example 13.5.1; Page 686,
head(EXA_C13_S05_01)
(med1351 = median(unlist(EXA_C13_S05_01), na.rm = TRUE)) # common median
addmargins(t1351 <- with(EXA_C13_S05_01, expr = {
  tmp <- cbind(Urban = table(URBAN < med1351), Rural = table(RURAL < med1351, useNA = 'no'))
  rownames(tmp) <- paste('Number of scores', c('above', 'below'), 'median')
  tmp
})) # Table 13.5.2
chisq.test(t1351, correct = FALSE)

# Example 13.6.1; Page 691,
head(EXA_C13_S06_01)
with(EXA_C13_S06_01, wilcox.test(X, Y, exact = FALSE, alternative = 'less'))

# Example 13.7.1; Page 699,
head(EXA_C13_S07_01)
ks.test(EXA_C13_S07_01$GLUCOSE, y = pnorm, mean = 80, sd = 6)

# Example 13.8.1; Page 705 (10th ed), Page 611 (11th ed)
(d1381 = data.frame(Air = c(12.22, 28.44, 28.13, 38.69, 54.91),
  Benzaldehyde = c(3.68, 4.05, 6.47, 21.12, 3.33),
  Acetaldehyde = c(54.36, 27.87, 66.81, 46.27, 30.19))) # Table 13.8.1
with(data = reshape2::melt(
```

```

    data = d1381,
    id.vars = NULL,
    value.name = 'Eosinophil',
    variable.name = 'Exposure'
  ), expr = kruskal.test(x = Eosinophil, g = Exposure))

# Example 13.8.2; Page 708 (10th ed), Page 613 (11th ed)
head(EXA_C13_S08_02)
with(data = reshape2::melt(
  data = EXA_C13_S08_02,
  id.vars = NULL,
  value.name = 'Value',
  variable.name = 'Hospital'
), expr = kruskal.test(x = Value, g = Hospital))

# Example 13.9.1; Page 713 (10th ed); Page 618 (11th ed)
head(EXA_C13_S09_01)
m1391 = as.matrix(EXA_C13_S09_01[LETTERS[1:3]], rownames.force = TRUE)
names(dimnames(m1391)) = c('Therapist', 'Model')
m1391 # Table 13.9.1
friedman.test(m1391)

# Example 13.9.2; Page 715 (10th ed); Page 620 (11th ed)
head(EXA_C13_S09_02)
m1392 = as.matrix(EXA_C13_S09_02[LETTERS[1:4]], rownames.force = TRUE)
names(dimnames(m1392)) = c('Animal', 'Dose')
m1392 # Table 13.9.2
friedman.test(m1392)

# Example 13.10.1; Page 720,
head(EXA_C13_S10_01)
with(EXA_C13_S10_01, cor.test(X, Y, method = 'spearman'))

# Example 13.10.2; Page 722,
head(EXA_C13_S10_02)
with(EXA_C13_S10_02, cor.test(V2, V3, method = 'spearman', exact = FALSE))

# Example 13.11.1-13.11.2; Page 728-729,
testosterone <- c(230, 175, 315, 290, 275, 150, 360, 425)
citricAcid <- c(421, 278, 618, 482, 465, 105, 550, 750)
robslopes::TheilSen(x = citricAcid, y = testosterone)
# textbook uses *central* median of ordered pairs, while ?robslopes::TheilSen uses *upper* median
summary(mblm::mblm(testosterone ~ citricAcid, repeated = FALSE)) # Figure 13.11.1 (11th ed)

```

Description

Examples for Chapter 14, *Survival Analysis*.

Value

No function defined for Chapter 14.

Examples

```
library(DanielBiostatistics10th)

# Example 14.3.1; Page 756 (10th ed), Page 652 (11th ed)
head(EXA_C14_S03_01)
head(d1431 <- within(EXA_C14_S03_01, expr = {
  TIME = .difftime(TIME, units = 'months')
  OS = survival::Surv(time = TIME, event = (VITAL != 'ned'))
}))
class(d1431$OS) # 'Surv'
head(d1431$OS)
summary(sf_1431 <- survival::survfit(OS ~ TUMOR, data = d1431)) # Table 14.3.2
plot(sf_1431, lty = 2:3, xlab = 'month', ylab = 'OS', main = 'Figure 14.3.1'); legend(
  x = 250, y = .9, legend = c('High', 'Low'), lty = 2:3)

# Example 14.4.1; Page 764 (10th ed), Page 659 (11th ed)
survival::survdiff(OS ~ TUMOR, data = d1431)

# Example 14.5.1; Page 769 (10th ed), Page 663 (11th ed)
head(EXA_C14_S05_01)
head(d1451 <- within(EXA_C14_S05_01, expr = {
  time = .difftime(time, units = 'weeks')
  PFS = survival::Surv(time, status)
  drug = relevel(structure(drug, levels = c('Opiate', 'Other'), class = 'factor'), ref = 'Other')
}))
summary(model1_1451 <- survival::coxph(PFS ~ drug + age, data = d1451))
# 'Opiate' has higher hazard compared to Drug='Other' (hazard ratio (HR) = 9.407, p < .001)
# With 'proportional hazard' assumption,
# ... we don't need to discuss Opiate vs. Other at any specific time point
confint(model1_1451)
# 'age' is not significant (p = .772), so it should be removed from the model
summary(model2_1451 <- survival::coxph(PFS ~ drug, data = d1451))
confint(model2_1451)
# 'Opiate' has higher hazard compared to Drug='Other' (hazard ratio (HR) = 9.923, p < .001)
plot(survival::survfit(PFS ~ drug, data = d1451), lty = 2:3,
  xlab = 'weeks', ylab = 'PFS', main = 'Figure 14.5.1'); legend(
  x = 35, y = .9, legend = c('Other', 'Opiate'), lty = 2:3)
```

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